

EXPERIMENTAL LARVAL INFESTATION OF ZEBU CATTLE AND ITS CROSSES: EFFECT OF ANTI-HISTAMINE ALONE OR IN COMBINED WITH DEXAME-THASONE TREATMENT ON THE ENGORGEMENT AND OVIPOSITION ON TICK FEEDING LESION

Mahmood Ameen Abdulla¹, Khairul Anuar Abdullah², Khalifa Sidik¹, Salmah Ismail¹, Mohammad Nazmul Hassan Mahziz¹, Suzainur Kulop Abdul Rahman¹ and Irwan Hamzah¹

¹Department of Allied Health Science, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia

²Department of Parasitology, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia

ABSTRACT: Four calves from each group of purebred Kedah kelantan (KK), halfbred KK X Friesian, and quarterbred KK X Friesian were experimentally infested with *Boophilus microplus* larvae. Two calves from each genotype were injected, intramuscularly with antihistamine while the remaining two animals in each genotype received the same dose of antihistamine and dexamethasone. Dexamethasone combined anti-histamine treatment suppress tick resistance as manifested by the production of higher number of engorged female ticks, higher mean weight of replete ticks, mean weight of eggs and mean number of larvae hatched from 1 g of eggs. In anti-histamine treated animals there was no reduction of resistant in all animals as manifested by a few ticks were able to feed successfully. At 1, 2, and 3 hours post-larval attachment in anti-histamine and dexamethasone treated cattle there was complete ablation of the cellular infiltration in the dermis beneath the tick mouthparts, especially eosinophil and basophils. There was little destruction of tissue. However, in anti-histamine treated cattle there were more cellular infiltration and degranulation in the dermis. The cells infiltrating the dermis were mainly eosinophils followed by neutrophs, mast cells and basophils and some of these cells showed sign of degranulation. At 24 hours post-larval attachment, animals treated with anti-histamine and dexamethasone showed reduction of, cellular infiltration, degranulation, size of the epidermal lesion and tissue damage. The neutrophils were the predominant cells within the epidermal lesions. However, animals in anti-histamine treatment showed edema, more cellular infiltration and degranulation, and destruction of tissues as compared to antihistamine and dexamethasone treated animals. In anti-histamine treated cattle the epidermal lesions were obviously larger and the percentage of eosinophils and basophils were higher than those of antihistamine and dexamethasone treated animals. (JUMMEC 2002; 2:135 - 141)

KEYWORDS: Kedah-Kelantan cattle, KK X Friesian cattle, *B. microplus*, dexamethasone, antihistamine, cellular response.

Introduction

The Zebus and its crosses are relatively resistant to the cattle tick, *B. microplus*, as compared to pure European type cattle (1,2,3). Many previous studies have shown that the acquisition of resistance to ticks has an immunological basis (4,5,6)

Previous studies have shown that the granulocytes especially eosinophils and basophils may play a major role in the rejection of ticks in the early stages of infestation (7,8,9). Brown et al., (7) used highly specific antisera

and demonstrate that basophils and to a lesser extent eosinophils, were responsible for the expression of resistance in guinea pigs infested with *A. americanum*. This finding strongly supported the effector role of these cells in the expression of immunity against ticks. Resistance of tick infestation is a tissue response (1,10,11). The ability of certain breeds of cattle to resist infesta-

Correspondence:

Mahmood Ameen Abdulla
Department of Allied Health Sciences, Faculty of Medicine
University of Malaya, 50603 Kuala Lumpur,
Malaysia

tion by ticks was greatly influenced by the ability of the skin to mount a cellular response.

The role of histamine in the expression of host resistance to ticks has been suggested (12, 13, 14, 15). Elevated histamine levels were found in blood (12) and skin (16) of cattle resistant to *B. microplus*. Histamine is a potent vasodilator, released at the site of injury (15, 17). It also increases permeability of the capillaries and thus serves to recruit blood cells to the area in response to injury, which may either be caused by trauma or invading organisms. It has been shown that histamine is chemotactic to eosinophils (18). The accumulation of eosinophils, among other cells, at the site of tick infestation may be a consequence of this effect (7). In most cases, the net effect is the manifestation of hypersensitivity reactions in the affected tissue (19)

The abrogation of acquired resistance to nematode infection in rodents and sheep by immunosuppressive glucocorticoids has been well documented (20, 21). In sheep this was associated with a reduction in the numbers of circulating leucocytes (21) and in rats, with reduced number of intestinal intraepithelial mast cells and eosinophils (20). Both cortisone and corticotropin treatments reduced the number of globule leucocytes in the rat mucosal tissue (22). Corticosteroid drugs have been widely used in several host species to modify both natural resistance and acquired immunity to nematode parasites. By their use it is possible to achieve maturity of parasites in otherwise unsuitable hosts (23), to increase natural susceptibility to infection (24), to suppress the development of immunity during a primary infection and to prevent the action of an established immune response upon a challenge infection (25).

The objectives of this study were to determine the effect of antihistamine alone and in combination with dexamethasone on, the engorgement and oviposition of

B. microplus, the cellular infiltration at tick-attachment sites in pure local Zebu cattle and its European crosses.

Material and methods

Ticks

Twenty days old *B. microplus* larvae were used for infestation. The adult female ticks were those collected from the University Putra Malaysia dairy herd (Friesian-Sahiwal) and the engorged adults, eggs and the larvae maintained in an incubator 28 °C at approximately 80% relative humidity. Two susceptible cattle were regularly used for the engorged tick production

Experimental animals

A total of 12 six-month old healthy calves that have been grazed in *B. microplus*-infested pasture were used in the study. The calves included four purebred Kedah-

Kelanatn (KK) cattle, four 50% KK and 50% Friesian, and four 25% KK and 75% Friesian cattle.

Infestation

The animals were infested and biopsies of tick feeding sites taken according to the technique described by Amin-Babjee and Riek (26). All twelve experimental steers were infested with 20,000 of *B. microplus* larvae each on the same day before the first injection of dexamethasone and antihistamine.

For infestation, larvae were placed along the vertebral column of each animal. Small batches of larvae were placed within aluminum rings cemented (Selly's adhesive) to the previously shaved areas of the skin of the lumber region of each animal. The larvae were confined within the rings by fine-meshed cloth secured with rubber bands. Biopsies of skin within the rings were taken at 1, 2, 3, and 24 hours after infestation using a 0.05 mm trephine, under local anesthesia (2 ml of 2% xylocaine).

Two animals from each genotype were injected, intramuscularly with 200 mg (25 mg/ml) of tripelemine hydrochloride (anti-histamine) daily for five days (Treatment 1) while the remaining two animals in each genotype received the same dose of the tripelemine hydrochloride and 16 mg (2 mg/ml) dexamethasone intramuscularly daily for five days (Treatment 2).

Collection and weighing of engorged female ticks

The numbers and weight of engorged females from each animal were recorded daily and the mean numbers and weight of all ticks engorged on individual animal was calculated according to the technique described by Amin-Babjee and Riek (26).

Measurement of weight of eggs

The masses of egg produced by engorged ticks from each animal were mixed, weighed in the same manner as described by Amin-Babjee and Riek (26)

Measurement of larval production from 1 g of eggs

The counting of larvae was done two weeks after hatching (active larvae) according to the technique described by Amin-Babjee and Riek (26).

Process of skin biopsies with larval attachment for light microscopy study. Skin biopsies with attached larvae were taken from each animal at 1, 2, 3, and 24 hours after larval infestations. Processing the skin biopsies from fixation, trimming, dehydration, clearing, infiltration, embedding and staining were as described by Amin-Babjee and Riek, (26).

Analysis of the cellular response

Serial sections containing attachment sites (cement substance plus mouthparts of the larvae) with maximum cellular reactions were selected and analyzed for cellular response. Eosinophils, neutrophils, basophils mononuclear cells and mast cells were counted using an oil immersion (1000x) objective in 0.15 mm² areas, covering the center of the lesion and areas adjacent on both sides. Analyses were performed on three different feeding sites for each animal. The results were expressed as the mean cell count.

Results

Tick biological parameters

Effect of antihistamine alone and combination of antihistamines and dexamethasone on the engorgement and oviposition of

B. microplus larvae in local Zebu and its crosses.

The mean numbers of engorged female ticks, their mean weights, the mean egg masses produced by each engorged female tick and the mean numbers of larvae produced per gram of eggs for each breed in antihistamine alone and in combined antihistamine and dexamethasone treatment groups are summarized in Table (1)

In general, the mean numbers of replete female ticks, mean weight of replete ticks, mean weight of egg masses and the mean numbers of larvae hatched from one gram eggs laid by replete female ticks dropped from antihistamine and dexamethasone treated breed were higher than those dropped from antihistamine alone.

Histology

Effect of antihistamine alone and combined antihistamine and dexamethasone on the cutaneous cellular responses at 1, 2, 3 and 24 hours post larval attachment sites in local Zebu and its crosses.

The temporal changes in cellular responses at tick attachment sites for the two treatments and three genotypes at 1, 2, 3, and 24 hours after larval infestation are summarized Table (2). Antihistamine and dexamethasone treatment completely ablated or reduced the cellular infiltration at tick attachment sites in all genotypes compare with antihistamine alone. Antihistamine treatment had little or no effect on the cellular infiltration.

Overall, various cell-type at tick-feeding lesions were higher in antihistamine than antihistamine and dexamethasone treatment in all genotypes (Plate 1 and 2; Table 2).

Table 1. Effect of Anti-Histamine Alone or Combined with Dexamethasone Treatment on the Engorgement and Oviposition Rates of *B. microplus* Following Experimental Infestation in Cattle

Genotype	Treatment	No. Engorg Tick	Wt. Engorg. Tick (mg)	Wt. egg/tick (mg)	No. Hatch Larvae/g
K K K K	1	3	136	73	8742
	2	28	139	74	9465
K K F F	1	78	152	77	9151
	2	265	156	79	10945
K F F F	1	172	155	77	10158
	2	435	157	80	10687

Treatment 1 = Antihistamine treatment;

Treatment 2 = Antihistamine and dexamethasone treatment

K K K K = Purebred Kedah-Kelantan (KK) cattle

K K F F = Crossbred (50% KK and 50% Friesian)

K F F F = Crossbred (25% KK and 75% Friesian)

For each treatment, the values are the average of two animals

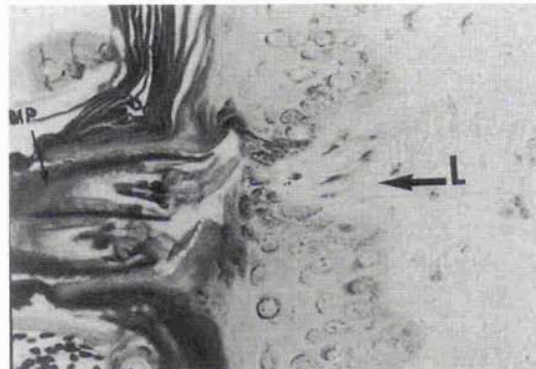


Plate 1. Feeding Sites of *Boophilus microplus* Larvae (3 h Postinfestation) in Anti Histamine and Dexamethasone Treated Zebu (K K K K) Cattle. The dermis is virtually ablated. (Stain with Alkaline Giemsa). As = Attachment Site; L- Few Cellular Infiltration (Primary Magnification x 40)

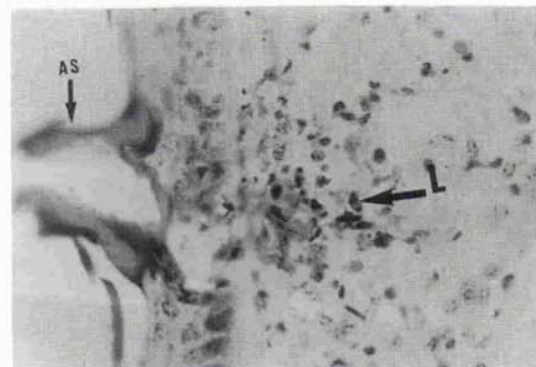


Plate 2. Feeding sites of *Boophilus microplus* Larvae (3 h postinfestation) in Anti-histamine treated Zebu (K K K K) Cattle. Note. The moderate infiltration of cells within the dermis. (stained with Alkaline Giemsa) As = Attachment site; L=Lesion (Primary Magnification x40)

In antihistamine treatment the number of eosinophils in purebred at 3, and 24 hours post-larval attachment were higher than the crossbred, while the number of basophils at 24 hours post-larval feeding sites were higher than the crossbred.

Discussion

The results of the current study showed that after an experimental infestation with 20,000 larvae of *B. microplus*, the mean numbers of replete female ticks, mean weight of replete ticks, mean weight of egg masses and the mean numbers of larvae hatched from one gram eggs laid by replete female ticks dropped from antihistamine and dexamethasone treated breed were higher than those dropped from antihistamine alone. Also the results of the present study indicated that the mean numbers of eosinophils and basophils infiltrating tick feeding sites were higher in antihistamine treated

than antihistamine and dexamethasone treated cattle at 1,2,3, and 24 hours post-larval attachment in all breeds. Results indicated that the injection of antihistamine did not affect the resistance level and the cellular infiltration in all three genotypes of cattle. It is possible that the intramuscular injection of antihistamine used in this study had no effect/influence on the migration and degranulation of eosinophils and basophils at attachment sites. However, several previous authors have reported that the use of antihistamine affects the expression of cattle resistance to ticks (14,16).

Antihistamine has some effect on suppression of resistance level in cattle. Riek (12) reported elevated levels of histamine in the blood of cattle resistant to *B. microplus* and stated that resistant hosts reacted more than susceptible hosts to intradermal injection of histamine. Tatchell and Bennett (27) stated that a significant increase occurred in the yield of engorged *B. microplus*

Table 2. Effect of anti-histamine alone or in combined with dexamethasone treatment on cutaneous cellular responses following experimental infestation with *B. microplus* larvae in cattle.

	Genotype Treatment	Hour	Mast cells	Eosinophils	Neutrophils	Mononuclear cells	Basophils
K K K K K	1	1	5.5	4.7	2.7	1.6	0.0
		2	6.5	8.8	5.8	1.5	2.5
		3	7.8	13.3	10.8	3.2	4.6
		24	0.0	9.3	274.3	3.3	10.4
	2	1	4.3	0.0	1.3	0.5	0.0
		2	4.5	0.0	2.7	0.8	0.0
		3	3.0	0.5	3.3	1.3	0.2
		24	0.0	2.0	235.0	3.3	2.2
K K K F F	1	1	7.8	3.2	3.2	2.3	0.0
		2	7.7	6.2	5.3	2.8	0.2
		3	6.8	7.7	8.3	2.0	4.5
		24	0.0	5.4	233.5	2.5	6.1
	2	1	4.3	0.0	0.5	1.7	0.0
		2	4.8	0.0	2.0	1.3	0.0
		3	4.8	0.0	2.2	2.2	0.0
		24	0.0	0.0	187.2	2.5	0.0
K F F F	1	1	6.7	3.5	2.2	1.8	0.0
		2	5.8	6.5	4.2	2.7	0.5
		3	7.5	6.2	6.7	2.2	1.3
		24	0.0	6.7	195.7	2.7	5.3
	2	1	4.8	0.0	0.2	0.5	0.0
		2	4.7	0.0	2.7	2.3	0.0
		3	3.2	0.0	3.3	1.8	0.0
		24	0.0	0.0	169.7	3.2	0.0

Treatment 1 Antihistamine; Treatment 2 Antihistamine and Dexamethasone; For each treatment, the values are the average of two animals.

from eight out of twelve cattle (*Bos taurus* and *Bos indicus*) treated with antihistamine promethazine hydrochloride. Histamine is released by mast cells in immediate hypersensitivity reactions in bovine (28) and its presence beneath the attachment sites of young larvae causes them to be detached (14). *Bos taurus* cattle with different degrees of resistance to *B. microplus* have responses to tick allergen that correlate with their resistance level. The total amount of histamine in the skin also correlates with both resistance and the immediate hypersensitivity reactions. Treatment with anti-histamine drug mepyramine maleate suppresses the cutaneous hypersensitivity reactions. The results suggest the main pharmacologically active agent in these reactions is histamine, and that the total amount of it available locally in the skin may have a role in the resistance to this parasite (16). Circumstantial evidence suggests that the earlier detachment of *B. microplus* larvae from highly resistant *Bos taurus* cattle follows the release of histamine at attachment sites. *In vivo* and *in vitro* experiments show that a proportion of the larvae will detach following injection or infusion of histamine. Other mediators such as bradykinin, prostaglandin E₂, 5-hydroxytryptamine and dopamine have little or no effect on tick behavior *in vivo*. Sensitivity to histamine declines as larval attachments stabilize, and repeated injections have no effect on the weight of the larvae after 3 days on the host (14).

Several authors have also suggested a role for histamine in the expression of resistance of laboratory animals (15, 17). Bagnall (13) showed that type-I histamine receptor antagonist, mepyramine, reduced the expression of tick resistance and obtained a slight decrease in the rejection of ticks when administered to guinea pigs resistant to *I. holocyclus* larvae. Allen (29) demonstrated that the host skin reaction to tick-resistant guinea pigs was characterized by an increase infiltration of basophils. The accumulation of basophils provided a localized high concentration of histamine. The histamine contained in the basophil granules can be released either by antibody dependent (30) or lymphokine mediated (31) immunological reactions, or by combination of both. Histamine at tick attachment sites may play a major role in the rejection of ticks. Basophils and mast cell granules are rich source of histamine (30). Thus the net consequence of these cellular interactions at tick feeding sites results in increased levels of histamine at tick feeding sites. Because of the chemical nature of histamine it could inhibit digestion or some other physiological process by possible altering enzyme structure or reactivity.

In other tick-host relationships, where infiltrating basophil leucocytes are more numerous in tick lesion (13,32) histamine may play a more important role in the rejection. Histamine has been shown to have a direct effect on tick attachment and feeding (14,15,17). Probably the most important action of histamine is in causing

irritation, leading to removal of ticks by self-grooming in highly resistant cattle (33). If histamine is responsible for repeated detachment of larvae during the first day of infestation, this could facilitate their removal by the host (34).

In this experiment, it was shown that only dexamethasone and not antihistamine, caused cellular ablation at tick attachment sites and this correlated with the suppression of tick resistance by the increase in numbers of ticks produced in the treated animals, this is consistent with the result of Eckblad et al (35) and Amin-Babjee et al., (36). Amin-Babjee et al (36) using European breeds cattle showed that dexamethasone, a potent glucocorticoid, when administered in a therapeutic dose induced immunosuppression, as shown by increase in the number of ticks, reduction in immediate hypersensitivity reactions, and obvious reduction or complete ablation of the cellular infiltration at the feeding sites of the tick larvae. In cattle, the administration of the glucocorticoid, a-fluoroprednisolone or dexamethasone is known to have a variety of potential effects: induction of lymphopenia (37), suppression of lymphocyte blastogenesis (38) and impaired of the functions of neutrophils (39).

Several workers have used immunosuppression drugs to study their effect on other host/parasite relationship. Rats infected with *S. ratti* and treated with corticosteroids do not expel their own burdens (20). Cortosone, as an anti-inflammatory agent, is known to affect the level of both eosinophils (40) and mast cells (41). Treatment of resistant sheep with immunosuppressive drug dexamethasone abolished resistance to challenge infections with *T. colubriformis* and was associated with both the depletion of globule leucocytes, mast cells and eosinophils from the small intestinal mucosa, as well as, a reduction in mucus larval migration inhibitory activity (42).

The present results showed that eosinophils and basophils are important cellular responses at feeding sites of larval stage and this response has contributed to resistance in the anti-histamine treated groups in all genotypes. Anti-histamine has no effect on the cellular infiltration. Both these cells were depleted at 3 and 24 hours after infestation in animal treated with anti-histamine and dexamethasone.

References

1. Panda DN, Ansari MZ, Sahai BN. and Jha GJ. Repeated infestations with larvae of *B. microplus* 2. Host reaction in crossbred calves. Indian J Anim Sci 1993; 63: 4-7.
2. Mangold AJ, Aguirre DH, Gaido AB and Gugliemone AA. Seasonal variation of ticks (Ixodidae) in *Bos taurus* X *Bos indicus* cattle under rotational grazing

- in forested and deforested habitats in northwestern Argentina. *Vet Parasitol* 1994; 54: 389-395.
3. Jonsson NN, Matschoss AL, Pepper P Green PE and Ansell J. Resistance of Holstein-Friesian cows to infestation by cattle tick (*B. microplus*). *Vet Parasitol* 2000; 89: 297-305.
 4. Brown SJ. Immunology of acquired resistance to tick. *Parasitol Today*; 1985; 1:165-171.
 5. Wikel SK and Whelen AC. Ixodid-host immune interaction. Identification and characterization of relevant antigens and tick-induced hoist immunosuppression. *Vet Parasitol* 1986; 20:149-174.
 6. Bechara GH, Morelli JR and Szabo MPJ. Skin test and tick immune status in susceptible and resistant cattle in Brazil. *Annals of the NY Academy of Sciences. Trop Vet Dis Control and prevention in the context of the new world order.* 2000; 916: 576-582.
 7. Brown SJ., Galli, SJ, Gleich GH and Askenase PW. Ablation of immunity to *A. americanum* by anti-basophil serum and eosinophils. in the expression of immunity to ectoparasites (ticks) in guinea pigs. *J Immunol* 1982; 129: 790-796.
 8. Gill HS. Kinetic of mast cell, basophil and eosinophils at *Hyalomma a. anatolicum* feeding sites on cattle and the acquisition of resistance. *Parasitol* 1986; 93: 305-315.
 9. Inokuma H, Kerlin RL, Kemp DH and Willadsen P. Effect of cattle tick (*B. microplus*) infestation on the bovine immune system. *Vet Parasitol* 1993; 47: 107-118.
 10. Fivaz BH, Nurton JP and Ptney TN. Resistance of restrained *Bos taurus* dairy bull calves to the bont tick *A. hebraeum*. *Vet Parasitol* 1991; 38: 299-315.
 11. Latif AA, Punyua DK, Walker AR and Fletcher JD. Histopathology of attachment sites of *A. variegatum* and *R. appendiculatus* on zebu cattle of varying resistance to ticks. *Vet Parasitol* 1991; 38: 205-213.
 12. Riek RF. Studies on the reactions of animals to infestation with ticks. VI. Resistance of cattle to infestation with tick *B. microplus*. *Aust J Agri Res* 1962; 13: 532-550.
 13. Bagnall BG. Cutaneous immunity to the tick *I holocyclus*. Ph D Thesis University of Sydney 1975; 186.
 14. Kemp DH and Bourne A. *B. microplus*: the effect of histamine on the attachment of cattle-tick larvae - studies *in vivo* and *in vitro*. *Parasitol* 1980; 80: 487-496.
 15. Wikel SK. Histamine content of tick attachment sites and the effect of H₁ and H₂ histamine antagonists on the expression of resistance. *Ann Trop Med Parasitol* 1982; 76: 179-185.
 16. Willadsen P, Wood GM and Riding GA. The relation between skin histamine concentration, histamine sensitivity and the resistance of cattle to the tick, *B. microplus*. *Z. Parasitkde* 1979; 59: 87-93.
 17. Paine SH, Kemp DH and Allen JR. *In vitro* feeding of *D. andersoni* : effect of histamine and other mediators. *Parasitol* 1983; 86: 419-428.
 18. Dale DC, Hubert CR and Fauci AS. Eosinophil kinetics in the hypereosinophilic syndrome. *J Lab Clin Med* 1976; 87: 487-490.
 19. Butterworth AE, Wassom GJ, Gleich DA and David JR. Damage to schistosoma mansonii induced directly by eosinophil major basic protein. *J Immunol* 1979; 122: 221-229
 20. Moqbel R and Denham DA. *S rattii*: the effect of betamethasone on the course of infection in rats. *Parasitol* 1978; 76: 289.
 21. Matthews DR, Brunton RV and Vlassoff A. Effect of dexamethasone on the ability of sheep to resist reinfestation with nematodes. *Vet Parasitol* 1979; 5: 65-72.
 22. Kent JF, Baker BL and Li CH. Effect of corticotropin and cortisone on globule leucocyte of rat. *Proc Soc Exp Bio. Med* 1954; 85: 636-637.
 23. Cross JH. The natural resistance of the white rat to *N. dubius* and the effect of cortisone on this resistance. *J Parasitol* 1963; 46: 175-185.
 24. Mathies AW. Certain aspects of the host parasite relationship of *A. tetraoptera* a mous pinworm. III. Effect of cortisone. *J Parasitol* 1962; 48: 244-248.
 25. Coker CM. Effect of cortisone in *T. spiralis* infections in non-immunized mice. *J Parasitol* 1955; 41: 498-504.
 26. Amin-Babjee SM and Riek RF. Histological changes in the skin following repeated infestations of cattle with *B. microplus*. *Kajian Veterinar* 1986; 18: 107-128.
 27. Tatchell RJ and Bennett GF. *B. microplus*: anti-histaminic and tranquilizing drugs and cattle resistance. *Exp Parasitol* 1969; 26: 369-377.
 28. Wells PW and Eyre P. Pharmacology of passive cutaneous anaphylaxis in the calf. *Cand J Phys Pharmacol* 1972; 50: 255-262.
 29. Allen JR. Tick resistance: basophils in skin reactions

- of resistant guinea pigs. *Inter J Parasitol* 1973; 3: 195-200
30. Askenase PW. Role of basophils, mast cells and vasoamines in hypersensitivity reactions with delayed time course. *Prog Allergy* 1977; 23: 199.
 31. Dvorak HF, Mihm MC and Colvin RB. Morphology of delayed type hypersensitivity reactions in man. I. Quantitative description of inflammatory response. *Lab Invest* 1974; 31: 111-130.
 32. Allen JR, Doube BM and Kemp DM. Histology of bovine skin reactions to *I. holocyclus* Neumann. *Can J Comp Med* 1977; 41: 26-35.
 33. Koudstaal D, Kemp DH and Kerr JD. *B. microplus*: rejection of larvae from British breed cattle. *Parasitol* 1978; 76: 379-386.
 34. Kemp DH, Koudastaal D, Roberts A and Kerr JD. *B. microplus*: the effect on host resistance on larval attachment and growth. *Parasitol* 1976; 73: 123-136.
 35. Eckblad WP, Stiller D, Woodard LF and Kuttler KL. Immune responses of calves antigenically stimulated and challenge exposed with *A. marginale* during tick infestation or treatment with dexamethasone. *Amer J Vet Res* 1984; 45: 431-436.
 36. Amin-Babjee SM, Willadsen P and Johnston LAY. Tick resistance, abrogation of the immune response to *B. microplus* by systemic injections of cortisone. *Malaysian Appl Biol* 1986; 15: 79-87.
 37. Wikie BN, Caoili F and Jacobs R. Bovine lymphocytes: Erythrocyte rosettes in normal lymphomatous and corticosteroid treated cattle. *Can J Comp Med* 1979; 43: 22-28.
 38. Muscoplat CC, Shope RE and Johnson DW. Effect of corticosteroids on responses of bovine peripheral blood lymphocytes cultured with PHA. *Am J Vet Res* 1975; 36: 1243-1244.
 39. Roth JA and Kaeberle ML. Effect of in vitro dexamethasone administration in bovine polymorphnuclear leucocyte function. *Infect Immunol* 1981; 33:434-441.
 40. Archer RK. The eosinophil response in the horse to intramedullary and intradermal injection of histamine, ACTH and cortisone. *J Pathol Bact* 1956; 72: 78.
 41. Cavallero C and Braccini C. Effect of cortisone on the mast cells of the rat. *Proc Soc Exp Biol Med* 1951; 87: 141.
 42. Douch PG, Harrison GB and Greer KS. Relationship of gastrointestinal histology and mucus antiparasite activity with the development of resistance to *Trichostrongylus* infections in sheep. *Vet Parasitol* 1986; 20: 315-331.